of the resulting amino ester nucleus, isolated as the tosylate salt (4), with thiopheneacetyl chloride provided the 3-cyanomethyl-7-thiopheneacetamido cephem ester (5a). Much decomposition occurred during ester cleavage of 3 or 5a with acid. In the case of 5a, however, some cephem acid material was isolated for antibacterial testing. The decomposition observed may result from carboxyl cyclization with the cyano moiety. The resulting iminolactone might then polymerize or otherwise lead to intractable material.

Table I provides gradient plate in vitro antibacterial activities for **5b** compared with other thiopheneacetamido cephem derivatives.

Experimental Section

Melting points were detd using a Mel-Temp app and are un-Uv spectra were detd in EtOH, ir spectra in CHCl₃ or as a cor. mull. Nmr spectra were obtd using a Varian HA-60 spectrometer in $CDCl_3$, acetone- d_6 , or $DMSO-d_6$. All cryst compds were characterized by ir, uv, nmr, and elemental anal. (C, H, N). Unless otherwise stated, anal. were within $\pm 0.4\%$ of the theor value.

tert-Butyl 3-Cyanomethyl-7-phenoxyacetamido-2-cephem-4carboxylate (2).—To a stirred, cooled soln contg 5.0 g (~ 10 mmoles) of bromide 1, dissolved in 90 ml of DMSO and 30 ml of DMF, was added solid Cu₂(CN)₂ (896 mg, 10 mmoles). After being stirred with slow warming to 20° over 3 hr, 10 ml of cold 5% HCl contg 4.0 g of FeCl₃ \cdot 6H₂O was added, and the mixt was stirred in the cold $(0-5^{\circ})$ for 15 min. C₆H₆ and satd aq NaCl were added, and the sepd org layer was washed with satd aq NaCl, satd NaHCO3, and satd NaCl and dried (MgSO4 with decolorizing charcoal). Filtn and evapn of the C_6H_6 soln yielded 4.0 g of crude 2 which was purified by column chromatog on silica gel-15% H2O. Compd 2 was eluted by 4% EtOAc in C₆H₆ and crystd from Et₂O, mp 117-120°.

tert-Butyl 3-Cyanomethyl-7-phenoxyacetamido-3-cephem-4carboxylate (3).—To a stirred, cooled soln contg 1.3 g (3 mmoles) of 2 dissolved in 40 ml of CH2Cl2 and 200 ml of i-PrOH was added dropwise 615 mg (3 mmoles) of m-ClC₆H₄CO₃H (85%) dissolved in 100 ml of *i*-PrOH and 100 ml of CH₂Cl₂. The mixt was stirred for 4 hr and allowed to warm slowly to room temp before being evapd to dryness. The residue was dissolved in 12 ml of 3:1 CH₃CN-DMF, and cooled in an ice bath before 1.3 g of SnCl₂ (anhyd) and 5 ml of AcCl were added. After standing 50 min in the cold and 45 min at room temp, the reaction mixt was evapd to dryness, and then was dissolved in C6H6; this soln was washed with cold 5% HCl, satd NaHCO₃, and satd NaCl, then dried (MgSO₄), and evapd to give 1.7 g of crude **3** as an oil that was purified by column chromatog. The desired material, eluted by 4-8% EtOAc in C₆H₆, did not crystallize, but was characterized by spectral data.

tert-Butyl 7-Amino-3-cyanomethyl-3-cephem-4-carboxylate Tosylate (4).-To a stirred soln contg 429 mg (1 mmole) of 3 dissolved in 20 ml of dry C₆H₆ was added 118 mg (1.5 equiv) of dry pyridine in 5 ml of dry $C_{6}H_{6}$; immediately following, 132 mg (1.5 equiv) of PCl₅ was added. After heating at 57° for 2 hr, the reaction mixt was cooled at room temp, evapd to dryness, and dissolved in 40 ml of cold MeOH. This soln was allowed to stand at room temp for 16 hr and then was evapd to dryness. To the residue was added 20 ml of THF and, after cooling, 20 ml of pH 4.5 buffer. After standing 20 min at room temp, THF was removed in vacuo, EtOAc was added to the residue, and the pH was adjusted to 6.5 by addn of NaHCO₃. The org layer was dried (MgSO₄) and evapd to give an oil. The cryst tosylate 4 mp 180-182°, was obtd by mixing EtOAc solns of the amine and TsOH.

tert-Butyl 3-Cyanomethyl-7-thiopheneacetamido-3-cephem-4carboxylate (5a).-To a stirred, cooled soln contg 467 mg (1.0 mmoles) of 4 suspended in 30 ml of Me₂CO was added 420 mg (5 mmoles) of solid NaHCO₃, followed by 482 mg (3 mmoles) of thiopheneacetyl chloride. The reaction mixt was stirred in the cold for 1 hr and at room temp for 3 hr; the Me₂CO was removed in vacuo, and the residue was dissolved in C_6H_6 . This soln was washed with cold 5% HCl, satd NaHCO3, and satd NaCl, and was dried (MgSO₄), evapd, and the residue was crystd from CCl_4 to give 418 mg, mp 164-166°.

C15H11F3O

В

3-Cyanomethyl-7-thiopheneacetamido-3-cephem-4-carboxylic acid (5b).-A soln of 455 mg of ester 5a in 40 ml of 98-100% $HCO_{2}H$ was stirred under N₂ for 2.5 hr at room temp. The HCO2H was removed in vacuo, and the residue was dissolved in EtOAc-NaHCO₃. The layers were sepd, and a second extn was performed with a NaHCO₃ soln. The aq exts were cooled, layered with EtOAc, and adjusted to pH 2.8 with 20% HCl. The org layer was washed with NaCl, dried (MgSO₄), and evapd to give 234 mg of a golden foam. Crystn from Et₂O gave 97 mg of acid 5b, mp 114-117°, characterized by spectral methods: uv, 268 m μ (ϵ 5400); ir (mull), C=O abs at 5.57, 5.80, and 6.03 μ . This material showed only 1 spot which was slightly faster moving than cephalothin on bioautogram (against Bacillus subtilis) in $MEK-H_2O$ (98:2).

Synthesis and Pharmacological Activity of Dialkylaminoalkyl Esters of Benzilic Acids **Containing Fluorine or Trifluoromethyl Groups**

I. LALEZARI,* M. HATEFI,

Department of Chemistry, Faculty of Pharmacy

M. A. KHOYI, N. GUITI, AND F. ABTAHI

Department of Experimental Medicine and Pharmacology, Faculty of Medicine, University of Tehran, Iran

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Continuing our studies on the synthesis of new potent local anesthetics,¹ and prompted by previous work on



166-168 (6.5)

Ia F

Ib

Ie н

 \mathbf{F}

F

н CF: 75

CFs 72198-200 (753) В C13H10F4O F н Id ^a All compds were analyzed for C, H, and the anal. values obtained were within $\pm 0.4\%$ of the calcd figures. All compds were also subjected to ir and nmr spectroscopy and showed the expected absorptions. ^b A. Fisher, B. A. Grigor, J. Packer, and J. Vaughan, J. Amer. Chem. Soc., 83, 4208 (1961), report mp ^c W. Funasaka, T. Ando, H. Ozahi, and K. Murakami, 111°. Yuki Gosei Kagaku Kyokai Shi, 17, 334 (1959) [Chem. Abstr., 53, 17970 (1959)], report mp 96-97°, bp 178-180° (15 mm). d N. Sharghi and I. Lalezari, J. Chem. Eng. Data, 10, 196 (1965), report bp 166-168° (5.5 mm).



^a See footnote a, Table I. ^b G. G. Smith and O. Larson, J. Amer. Chem. Soc., 82, 104 (1960), report mp 62-63°. Smith and Larson^b report mp 121.5-122.5°.

(1) N. Sharghi, I. Lalezari, G. Niloofari, and H. Golgolab, J. Med. Chem., 12, 696 (1969).



TABLE III

No.	\mathbf{R}_{1}	\mathbf{R}_2	Rs	R	Yield, %	Mp, °C	Formula ^a
IIIa	\mathbf{F}	н	н	Н	618	116	$C_{14}H_{11}FO_3$
IIIb	\mathbf{F}	\mathbf{F}	н	Н	69	143	$\mathrm{C_{14}H_{10}F_2O_3}$
IIIc	Н	н	CF_3	H	65	102	$C_{15}H_{11}F_{3}O_{3}$
IIId	\mathbf{F}	н	CF_3	H	72	122	$C_{15}H_{10}F_4O_8$
IVa	\mathbf{F}	н	н	$(CH_2)_2N(CH_3)_2 \cdot HCl$	54	192	$C_{18}H_{21}ClFNO_{3}$
IVb	\mathbf{F}	\mathbf{F}	н	$(CH_2)_2N(CH_3)_2\cdot HCl$	51	188	$\mathrm{C_{18}H_{20}ClF_2NO_3}$
IVc	\mathbf{F}	н	н	$(CH_2)_3N(CH_3)_2 \cdot HCl$	64	184	$\mathrm{C}_{19}\mathrm{H}_{23}\mathrm{ClFNO}_3$
IVd	\mathbf{F}	\mathbf{F}	H	$(CH_2)_3N(CH_3)_2 \cdot HCl$	62	178	$\mathrm{C}_{19}\mathrm{H}_{22}\mathrm{ClF}_{2}\mathrm{NO}_{3}$
IVe	н	н	CF_3	$(CH_2)_2N(CH_3)_2 \cdot HCl$	66	177	$\mathrm{C}_{19}\mathrm{H}_{21}\mathrm{ClF_3NO_3}$
IVf	\mathbf{F}	н	н	$(CH_2)_2 N (C_2H_5)_2 \cdot HCl$	64	170	$\mathrm{C}_{20}\mathrm{H}_{25}\mathrm{ClFNO}_3$
IVg	\mathbf{F}	\mathbf{F}	н	$(CH_2)_2 N (C_2H_5)_2 \cdot HCl$	52	182	$\mathrm{C}_{20}\mathrm{H}_{24}\mathrm{ClF}_2\mathrm{NO}_3$
IVh	\mathbf{F}	н	\mathbf{H}	$(CH_2)_2 N (CH_2)_5 \cdot HCl$	56	170	$\mathrm{C_{21}H_{25}ClFNO_{3}}$
IVi	\mathbf{F}	\mathbf{F}	H	$(\mathrm{CH}_2)_2\mathrm{N}(\mathrm{CH}_2)_5\!\cdot\!\mathrm{HCl}$	60	181	$\mathrm{C}_{21}\mathrm{H}_{24}\mathrm{ClF_2NO_3}$
IVj	н	н	\mathbf{CF}_{3}	$(\mathbf{CH}_2)_2 \mathbf{N} (\mathbf{C}_2 \mathbf{H}_5)_2 \cdot \mathbf{HCl}$	50	165	$C_{21}H_{25}ClF_3NO_3$
IVk	F	н	\mathbf{CF}_3	$(CH_2)_2 N (C_2H_5)_2 \cdot HCl$	52	158	$\mathrm{C}_{21}\mathrm{H}_{24}\mathrm{ClF_4NO_3}$
IVl	\mathbf{F}	н	\mathbf{CF}_{3}	$(CH_2)_2 N (CH_2)_5 \cdot HCl$	53	165	$\mathrm{C}_{22}\mathrm{H}_{24}\mathrm{ClF_4NO_3}$
IVm	н	н	\mathbf{CF}_{3}	$(CH_2)_2N(CH_2)_5 \cdot HCl$	80	159	$\mathrm{C}_{22}\mathrm{H}_{25}\mathrm{ClF_3NO_3}$
IVn	\mathbf{F}	н	н	$(CH_2)_2 N (CH_2 C_6 H_5)_2 \cdot HCl$	60	186	$C_{30}H_{29}ClFNO_3$
IVo	\mathbf{F}	\mathbf{F}	н	$(CH_2)_2 N (CH_2 C_6 H_5)_2 \cdot HC^2$	56	172	$\mathrm{C}_{30}\mathrm{H}_{28}\mathrm{ClF}_2\mathrm{NO}_3$
IVp	н	\mathbf{H}	\mathbf{CF}_{3}	$(CH_2)_2 N (CH_2 C_6 H_5)_2 \cdot HCl$	72	169	$C_{31}H_{29}ClF_3NO_3$
IVq	F	н	CF_3	$(\mathrm{CH}_2)_2\mathrm{N}(\mathrm{CH}_2\mathrm{C}_6\mathrm{H}_5)_2\cdot\mathrm{HCl}$	54	162	$\mathrm{C_{31}H_{28}ClF_4NO_3}$
IVr	н	н	CF_3	$(\mathrm{CH}_2)_3\mathrm{N}(\mathrm{CH}_3)_2\cdot\mathrm{HCl}$	65	140	$\mathrm{C}_{20}\mathrm{H}_{23}\mathrm{ClF_3NO_3}$
a		1					

^a See footnote a Table I. ^b See footnote b, Table II, mp 113-114°.

organofluorine drugs, we have prepared the title compounds. Several dialkylaminoalkyl esters of fluorinated and trifluoromethylated benzilic acids have been prepared and subjected to pharmacological screening. Some of the compounds were found to be potent local anesthetics, and to have anticholinergic and antihistaminic activities. **Pharmacological Evaluation.**—Compds IVb, IVd, IVe, IVh, IVi, IVk, IVm, and IVr were screened for surface anesthetic, anticholinergic, and antihistaminic activity. The results are summarized in Table IV. These compds were injected intraspinally, in a concn of 0.5%, in rabbits. The duration of urethral areflexia²

The general synthesis is shown in Scheme I.

(2) R. A. Turner, "Screening Methods in Pharmacology," Academic Press, New York, N. Y., 1965, p 268.

		Rabbit cornea		Guinea pig cornea-				
Compd	Concn, %	Potency	Duration, min	Potency	Duration, min	Dose. µg∕ml	Anti- cholinergic	Anti- histaminio
IVb	1	0	0	0	0	$\frac{1}{10}$	70 100	$\frac{57}{100}$
IVd	1	0	0	0	0	1 10	$\frac{5}{43}$	$\frac{10}{52}$
IVe	$\begin{array}{c} 0.25 \\ 0.50 \\ 1 \end{array}$	$\begin{array}{c} 0.02 \ (0{-}0.04) \\ 0.72 \ (0.64{-}0.81) \\ 0.93 \ (0.88{-}0.98) \end{array}$	0-3 12-18 17-21	$\begin{array}{c} 0.25 \ (0.17 0.33) \\ 0.88 \ (0.82 0.94) \\ 1.00 \end{array}$	2-7 12-24 18-51	$1 \\ 10$	$\frac{52}{98}$	$\frac{68}{100}$
IVh	1	0.45 (0.36-0.55)	4–17	0.82(0.75 - 0.90)	0-36	$\begin{array}{c} 1 \\ 10 \end{array}$	55 100	$\frac{85}{100}$
IVi	1	0.56(0.47 - 0.66)	8–16	0.80 (0.73-0.88)	5-54	$\begin{array}{c} 1 \\ 10 \end{array}$	58 96	$\frac{52}{100}$
IVk	$\begin{array}{c} 0.25 \\ 0.50 \\ 1 \end{array}$	$\begin{array}{c} 0.05 \ (0 - 0.09) \\ 0.55 \ (0.45 - 0.64) \\ 1.00 \end{array}$	0-2 6-18 21-33	$\begin{array}{c} 0.33 \ (0.24 0.42) \\ 0.97 \ (0.94 1.00) \\ 1.00 \end{array}$	2-11 21-36 36-96	$1 \\ 10$	$\frac{15}{100}$	27 100
IVm	$egin{array}{c} 0.25 \ 0.50 \ 1 \end{array}$	$\begin{array}{c} 0.10 \; (0.040.16) \\ 0.82 \; (0.750.88) \\ 1.00 \end{array}$	0-8 13-21 30-48	$\begin{array}{c} 0.50 \; (0.400.60) \\ 0.99 \; (0.971.00) \\ 1.00 \end{array}$	$\begin{array}{c} 4-14 \\ 18-42 \\ 27-84 \end{array}$	1 10	$\frac{23}{97}$	$\frac{50}{100}$
IVr	$\begin{array}{c} 0.25 \\ 0.50 \end{array}$	0.98 (0.95–1.00) 1.00	$26 - 39 \\ 30 - 55$	$0.48(0.38-0.58) \\ 0.98(0.95-1.00)$	4-17 30-40	$\frac{1}{10}$	$\frac{25}{83}$	$\frac{10}{57}$
Benactyzine	$\begin{array}{c} 0.25 \\ 0.50 \\ 1 \end{array}$	$\begin{array}{c} 0.40 \ (0.300.49) \\ 0.59 \ (0.500.69) \\ 0.98 \ (0.951.00) \end{array}$	5-16 7-23 19-35	$\begin{array}{c} 0.39 \ (0.29 0.48) \\ 0.46 \ (0.37 0.56) \\ 0.99 \ (0.97 1.00) \end{array}$	5-11 7-26 23-40	$\begin{array}{c} 0.1 \\ 1 \end{array}$	100 100	$\frac{21}{87}$
Cocaine	$\begin{array}{c} 0.25 \\ 0.50 \\ 1 \end{array}$	$\begin{array}{c} 0.13 \ (0.080.18) \\ 0.54 \ (0.470.62) \\ 0.95 \ (0.920.98) \end{array}$	$2-6 \\ 7-15 \\ 16-24$	$\begin{array}{c} 0.09 \ (0.04 0.15) \\ 0.55 \ (0.45 0.64) \\ 0.61 \ (0.52 0.70) \end{array}$	$0-5 \\ 4-18 \\ 8-21$			

TABLE IV^a

^a Surface anesthesia was tested according to the method of M. R. A. Chance and H. J. Lobstein, *J. Pharmacol. Exp. Ther.*, **82**, 203 (1944); and the anesthetic potency was calculated for the first 18 min, A. H. Campbell, J. A. Stasse, G. H. Lord, and J. E. Willson, *J. Pharm. Sci.*, **57**, 2045 (1968). A potency of 1.00 indicates an ouset of anesthesia in 1 min and a duration of at least 18 min. Reduction of contractions (%) against acetylcholine, $0.1 \mu g/ml$, and histamine dihydrochloride, $0.01 \mu g/ml$. Assay was carried out on isolated organs (ref 2, p 43). The test compounds were added to the bath 30-60 sec before the agonists.

was 3, 0, 19, 15, 16, 34, 20, and 23 min, resp. Benacty $zine \cdot HCl$ and $procaine \cdot HCl$, in the same concn, caused 22 and 5 min of areflexia, resp. In guinea pigs, IVe, IVk, IVm, and IVr caused conjunctival congestion in the first hr and slight opalescence of the cornea was present 48 hr after instillation. The LD_{50} of IVm in mice, estimated by the moving average method³ was 121 (85-172) mg/kg. Substitution in the Ph rings of benzilic acid β -diethylaminoethyl ester HCl (benactyzine HCl) resulted in decreased anticholinergic activity but did not affect the antihistaminic activity. The antihistaminic activity was decreased when the dialkylaminoethyl chain was replaced by dialkylaminopropyl. CF_3 substituted in the Ph rings increase the surface anesthetic activity. Compds bearing Et_2N or piperidino groups, compared with those having Me₂N group, were found to be more potent surface anesthetics.

Experimental Section⁴

3-Trifluoromethylphenyl-4'-fluorobenzyl Ketone (Id.)—m-Trifluoromethylbenzonitrile (20.5 g) (0.12 mole) in 48 ml of dry Et₂O was added dropwise to a Grignard reagent prepared from 22 g (0.15 mole) of *p*-fluorobenzyl chloride and 3.9 g (0.162 g-atom) of Mg. The reaction mixt was stirred, first at room temp and then on a steam bath for 3 hr, and decompd with ice and HCl. The ketone was extd with Et_2O , washed, dried, and distd after removal of the solvent.

Compd Ic was prepared similarly. Ia and Ib were obtd by the method described in refs given in Table I, footnotes b-d.

3-Trifluoromethyl-4'-fluorobenzil (IId).—SeO₂ (4.4 g, 0.04 mole) was dissolved in 50 ml of glacial AcOH. To the hot soln was added 11.2 g (0.04 mole) of Id and the mixt was refluxed for 3 hr. The Se ppt was filtered off and the soln was dild with H₂O. After cooling, the crystals were collected and recrystd from dil EtOH. The other compds II were prepd similarly.

3-Trifluoromethyl-4'-fluorobenzilic Acid (IIId).—To a soln prepd from 25 g of KOH and 100 ml of 50% EtOH was added 29.6 g of IId and the mixt was refluxed for 3 hr and concd on a steam bath. The residue was dild with H_2O and extd with Et_2O to remove the impurities. The clear H_2O soln was decolorized by charcoal, filtered, and acidified. The oily ppt was extd with Et_2O , and the solvent was evapd. The substituted benzilic acid was crystd slowly and was recrystd from dil EtOH. The other compds III were obtained similarly.

Dialkylaminoalkyl Esters (IV).—The appropriate benzilic acid (0.05 mole) and dialkylaminoalkyl chloride (0.05 mole) in 15 ml of dry *i*-PrOH was refluxed for 24 hr and the solvent was removed under reduced pressure. The residue was dissolved in 20 ml of H₂O, made alk with coucd K₂CO₃ soln and extd with Et₂O. The Et₂O ext was dried (MgSO₄) and filtered. Dry HCI was bubbled into the resulting soln until the ppt was complete (normally the mixt was stored 1 day in the ice box to complete the crystn). The esters were recrystd from EtOH.

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⁽³⁾ C. S. Weil, Biometrics, 8, 249 (1952).

⁽⁴⁾ Melting points were taken on a Kofler hot stage microscope. The ir spectra were detd with a Leitz Model III spectrograph. Nmr spectra were obtd on a Varian A60A instrument.